

5 α -CHOLESTANE-3 β ,6 β ,15 α ,16 β ,26-PENTOL: A POLYHYDROXYLATED STEROL
 FROM THE STARFISH *HACELIA ATTENUATA*[†]

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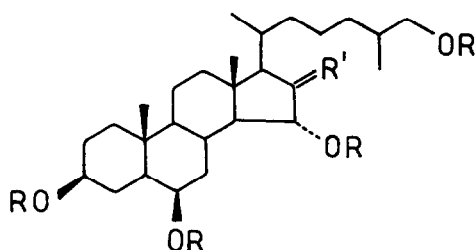
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Abstract. - A pentahydroxylated sterol isolated from a starfish, *Hacelia attenuata*, has been shown by spectral data and chemical transformations to be 5 α -cholestane-3 β ,6 β ,15 α ,16 β ,26-pentol.

Polyhydroxysterols are not uncommon to marine species and they have been observed especially in soft corals and gorgonians¹. Although starfish contain a number of polyhydroxysterols as aglycone constituents of saponins², free hydroxylated sterols have never been encountered before in this class of marine animals. In our continuing search for active saponins in starfish we have now isolated and characterized a novel polyhydroxylated sterol, 5 α -cholestane-3 β ,6 β ,15 α ,16 β ,26-pentol (1), from the mediterranean starfish *Hacelia attenuata*. The material was obtained in 0.004% yield (dry weight basis) from the chloroform-methanol 9:1 extract of the liophylized "starfish" by preparative h.p.l.c. (LC/system 500 Waters; column: prep. Pak 500 SiO₂; solvent. chloroform-methanol from 9:1 to 8:2) followed by chromatography on Sephadex LH-20 in MeOH and eventually reversed phase h.p.l.c. (C₁₈ μ -bondapak, MeOH-H₂O, 65:35).

The sterol (1), m.p. 197-199°C, $[\alpha]_D^{25} \pm 0^\circ$, had molecular formula C₂₇H₄₈O₅ as deduced from the mass spectrum (M⁺/e 452) and single-frequency off-resonance decoupled ¹³C-n.m.r. Acetylation with acetic anhydride and pyridine at room temperature led to the introduction of four acetyl groups 2, strong deacylated peaks at m/e 560 (M⁺-CH₃CO₂H), 500 (M⁺-2 CH₃CO₂H), 440 (M⁺-3CH₃CO₂H) and 380 (M⁺-4 CH₃CO₂H), 4 $\underline{\text{CH}_3\text{-C=O}}$ at δ 2.02, 2.05, 2.06 and 2.08. Oxidation of



- 1 R = H, R' =
- 2 R = Ac, R' =
- 3 R = Ac, R' = O

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the tetraacetate **2** using Jones reagent gave the ketone **3**, M^+/e 618. The single-frequency off-resonance decoupled ^{13}C -n.m.r. spectrum of **1** showed three doublets at 84.7, 82.2 and 71.3 (in the spectrum of the tetraacetate **2** the latter signal was splitted in two doublets at 72.8 and 73.3 ppm) and one triplet at 67.4 ppm establishing the presence of four secondary and one primary hydroxyls in **1**. The ^1H -n.m.r. spectrum of **1** (Table I) showed only two methyl doublets, centered at δ 0.93 and 0.99, which were transformed into singlets on irradiation at δ 1.60 and 1.96, respectively, thus locating the primary hydroxyl function at C-26. The 7-lines multiplet ($W_{\frac{1}{2}} = 20$ Hz) at δ 3.57 in the ^1H -n.m.r. spectrum of **1** is typical of the 3α -proton of an A/B *trans* steroid, while the quite narrow nature of the signal at δ 3.76 (in the spectrum of **1** this signal is superimposed on the signal for H-15, but in the spectrum of the tetraacetate **2** it is observed as well separated broad singlet at δ 4.98 with $W_{\frac{1}{2}} = 7$ Hz) was suggestive of the equatorial proton associated with the 6β -hydroxyl group³. The $3\beta,6\beta$ -dihydroxy oxidation pattern in **1** was supported from the ^{13}C -n.m.r. chemical shifts of the carbons 1-12 in **1** (Table II) which are in close agreement with those of the corresponding atoms in 5α -cholestane- $3\beta,6\beta$ -diol⁴. Several features of the ^1H -n.m.r. spectrum of **1** suggested that the two remaining secondary hydroxyls were located on the ring D. The 270-Mz n.m.r. displayed two doublets of doublets at δ 4.00 (H-16 $J_1 = 8.5$ Hz, $J_2 = 3$ Hz) and 3.80 (H-15 $J_1 = 10.5$ Hz, $J_2 = 3$ Hz) coupled each to other (3 Hz) as confirmed by decoupling. The magnitude of the coupling constant between H_{15} and H_{16} is only compatible with a *trans* relationship and the whole pattern is consistent with a $15\beta,16\alpha$ -protons system on a 17α -H C/D *trans*-steroid bearing 15α - and 16β -hydroxy groups. Further evidence was provided by several spectral features of the ketone **3**: a) the mass spectrum⁵ displayed two diagnostically important ions at m/e 448 (a) and 433 (b), namely side chain loss with migration of one hydrogen from the side chain to the nucleus and 18-methyl fission, typical of 16-ketosteroids⁶; b) the ^1H -n.m.r. displayed a doublet ($J = 14$ Hz) at δ 4.91 which had to be associated with the 15β -proton of a 14α -steroid; c) the transformation of the tetraacetate **2** into the ketone **3** was accompanied by change in the resonance frequency of the CH_3 -18 protons (δ 1.01 \rightarrow 0.96) which is only compatible with a 16β -oriented hydroxy group in **2**⁹.

The $3\beta,6\beta$ -hydroxylation pattern has been encountered before among polar marine steroids and recently in steroidal aglycones of cyclic saponins from two starfishes⁷, the functionalization of the CH_3 -26 is reminiscent of the bile alcohols, myximol and deoxymyximol, isolated from rayfish⁸, but the $15,16$ -diol structure is an unprecedented feature among sterols reported earlier.

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TABLE I - 270 MHz ^1H -n.m.r. data in δ (Hz)

Compound	$\text{CH}_3\text{-18}^{\text{a}}$	$\text{CH}_3\text{-19}^{\text{a}}$	$\text{CH}_3\text{-21}$	$\text{CH}_3\text{-27}$	H-3	H-6	H-15	H-16	$\text{H}_2\text{-26}$
1(CD_3OD)	0.95	1.07	0.99 (d 7)	0.93 (d 7)	3.57 (7-lines $W_{\frac{1}{2}}$ 20)	3.76 ^b	3.80 ^c (d 10.5,3)	4.00 (dd 8.5,3)	3.46 ^d (dd 12,6)
2(CDCl_3)	1.01	1.04	0.93 (d 7)	0.91 (d 7)	4.72 (7-lines $W_{\frac{1}{2}}$ 20)	4.98 (bs, $W_{\frac{1}{2}}$ 7)	4.45 (dd 10.5,2.5)	3.92 ^e	3.94-3.82 (dd 12,7) (dd 12,8)
3(CDCl_3)	0.96	1.05	1.01 (d 7)	0.92 (d 7)	4.72 (7-lines $W_{\frac{1}{2}}$ 20)	5.01 (bs, $W_{\frac{1}{2}}$ 7)	4.91 (d 14)	-	3.95-3.84 (dd 12,7) (dd, 12,8)

a.- Calculated according to Zurcher⁹ : 1, $\text{CH}_3\text{-18}$:0.97, $\text{CH}_3\text{-19}$ 1.05, 2, $\text{CH}_3\text{-18}$:1.02, $\text{CH}_3\text{-19}$:1.06, 3, $\text{CH}_3\text{-18}$:0.94, $\text{CH}_3\text{-19}$:1.08.

b.- Partially overlapped with the H-15 signal.

c.- The 10.5 Hz coupling between H_{15} and H_{14} was proved by decoupling.

d.- The highfield part of the AB portion of the $\text{HC-CH}_2\text{-OH}$ ABX system is submerged under methanol signal

e.- Overlapped with the H-26 signal.

TABLE II - Carbon-13 chemical shifts (ppm, TMS=0)^a

Compound	1	2	3	4	5	6	7	8	9	10	11	12	13	14
1(py- d_5)	39.2	32.5	71.3	37.0	48.3	71.3	41.1 ^b	30.9	55.0	36.1	21.3	41.2 ^b	43.9	61.0
1(CD_3OD)	39.8	32.2	72.5	36.4	c	75.2	41.9	31.2	55.8	36.6	21.9	40.6	44.7	61.1
2(CDCl_3)	38.0	27.2	73.3	30.9	46.0	72.8	36.0	30.3	53.6	35.4	20.6	40.1	43.3	56.2
	15	16	17	18	19	20	21	22	23	24	25	26	27	
1(py)	84.7	82.2	59.4	15.0	16.2	30.2	18.4	36.8	24.3	34.4	36.6	67.4	17.3	
1(CD_3OD)	85.0	82.9	59.9	15.0	16.3	30.9	18.6	37.4	24.8	34.9	37.0	68.4	17.3	
2(CDCl_3)	88.4	79.3	58.7	14.5	15.2	29.4	17.9	35.8	23.6	33.7	32.4	69.5	16.9	

a.- The carbon signals were assigned by means of single-frequency off-resonance decoupling techniques, by using the hydroxy substituent effects found in simpler steroids¹⁰ and the expected deviations from additivity for proximate diols¹¹, from comparison with the published data on the 5α -cholestane-3 β ,6 β -diol resonances and from comparison of the spectrum of the pentol 1 with that of its tetraacetate 2

b - Assignment may be reserved.

c.- Signal under solvent signal.

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- 5.- m.s. of compound 3: m/e (%) 618 (M^+ < 1%), 558 (5), 543 (10), 498 (8), 483 (12), 448 (a , 2), 438 (20), 433 (b , 34), 423 (20), 388 (a -CH₃CO₂H, 12), 373 (b -CH₃CO₂H, 34), 328 (a -2CH₃CO₂H, 70), 313 (b -2CH₃CO₂H, 50), 295 (18), 268 (a -3CH₃CO₂H, 100), 253 (b -3CH₃CO₂H, 70).
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